

5. Paris, M., Boucher, F. et Cosson, L. (1975) *Pl. Med. et Phytoth.* **IX**, 136.
6. Paris, M., Ghirlanda, C., Chaigneau, M. et Giry, L. (1973) *C.R. Ac. Sc. Paris* **276**, 205.
7. Boucher, F. (1972) *Mémoire du DEPS de Chimie des Médicaments naturels*. Paris.
8. Doorenbos, N. J., Fetterman, P. S., Quimby, M. W. et Turner, C. E. (1971) *Ann. N.Y. Acad. Sci.* **191**, 3.
9. Small, E., Beckstead, H. et Chan, A. (1975) *Economic Botany* **29**, 219.
10. Bassaz, F. A., Dusek, D., Seigler, M. S. et Haney, A. W. (1975) *Biochem. Systemat. Ecol.* **3**, 15.
11. Boucher, F. (1976) *Mémoire de D.E.A. de Physiologie Végétale Approfondie*. Paris.
12. Haney, A. et Kitscheid, B. B. (1973) *Econ. Botany* **27**, 193.

*Phytochemistry*, 1977, Vol. 16, pp. 1448–1450. Pergamon Press. Printed in England.

## TWO NEW 4 $\alpha$ -METHYLSTEROLS IN THE SEEDS OF *BRASSICA NAPUS*

T. ITOH, K. UCHIKAWA, T. TAMURA and T. MATSUMOTO

College of Science and Technology, Nihon University, 8, Kanda Surugadai, 1-chome, Chiyoda-ku, Tokyo, 101 Japan

(Received 31 March 1977)

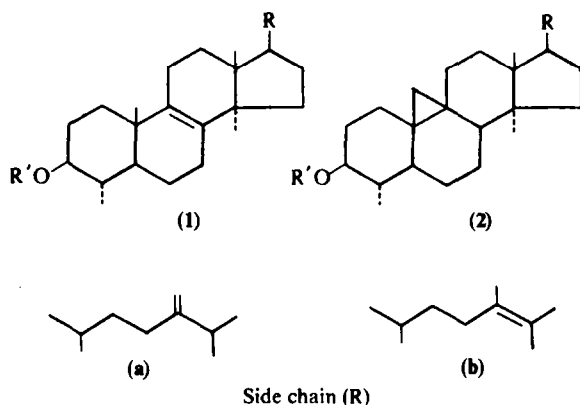
**Key Word Index**—*Brassica napus*; Cruciferae; 4 $\alpha$ -methylsterols; 4 $\alpha$ ,14 $\alpha$ ,24-trimethylcholesta-8,24-dien-3 $\beta$ -ol; 4 $\alpha$ ,14 $\alpha$ ,24-trimethyl-9 $\beta$ ,19-cyclocholest-24-en-3 $\beta$ -ol; seeds.

Our previous paper described the occurrence of 24-methylenelanost-8-en-3 $\beta$ -ol in the 4,4-dimethylsterol fraction from the seeds of *Brassica napus* [1]. We now report the detection and tentative identification of two new sterols in the 4-monomethylsterol fraction separated from the seeds. 4 $\alpha$ -Methylsterols hitherto found in the 4-monomethylsterol fraction from the Cruciferous seeds are 31-norlanostenol and/or 4 $\alpha$ -methylzymostenol [2], lophenol [3], 31-norlanosterol [2], obtusifoliol [2, 4], gramisterol (24-methylenelophenol) [2, 4] and citrostadienol [2, 4, 5].

The 4-monomethylsterol fraction separated from the unsaponifiable matter of the seeds of *B. napus* [1] was acetylated with Ac<sub>2</sub>O–Py. The acetate fraction (108 mg) obtained was separated into six bands by PLC on AgNO<sub>3</sub>–Si gel (1:4) with two developments using CCl<sub>4</sub>–CH<sub>2</sub>Cl<sub>2</sub> (5:1). The fraction (3 mg) recovered from the second band (*R<sub>f</sub>* 0.47) from the solvent front gave two peaks on GLC. GC–MS of the faster eluted major component (A, 68%) showed that it was an acetate of a C<sub>30</sub> sterol with two double bonds (*m/e* 468, M<sup>+</sup>, C<sub>32</sub>H<sub>52</sub>O<sub>2</sub>) of which one was present in the side chain (*m/e* 341, M<sup>+</sup> – C<sub>9</sub>H<sub>17</sub> [SC] – 2H). The fragment ions at *m/e* 301 (M<sup>+</sup> – SC – C<sub>3</sub>H<sub>6</sub>)

and 287 (*m/e* 301 – CH<sub>2</sub>) indicated the presence of an additional C-32 methyl group in the ring system. Furthermore, the fragment ions at *m/e* 384 (M<sup>+</sup> – C<sub>6</sub>H<sub>12</sub>) and 369 (*m/e* 384 – Me) showed that the side chain double bond was located either at C-24(28) or at C-24(25) [6]. The *RR*, of the steryl acetate was 1.82 (SE-30), 1.74 (Dexil-300), 1.76 (OV-17) and 1.79 (OV-210). Since the data from MS, GLC and argentation TLC were identical with those of authentic 4 $\alpha$ ,14 $\alpha$ ,24-trimethyl-5 $\alpha$ -cholesta-8,24-dien-3 $\beta$ -yl acetate (1b, R' = Ac) prepared from obtusifoliyl acetate (1a, R' = Ac), the steryl acetate A is considered to have the structure 1b, R' = Ac.

GC–MS of the slower eluted minor component (B, 32%) showed that it was also an acetate of a C<sub>30</sub> sterol with two double bonds (*m/e* 468, M<sup>+</sup>). The fragment ions at *m/e* 341, 384 and 369 and at *m/e* 301 and 287 indicated the presence of a double bond located at C-24(28) or at C-24(25) in the side chain and an additional C-32 methyl group in the ring system, respectively. Furthermore, the ion at *m/e* 285 (M<sup>+</sup> – C<sub>10</sub>H<sub>16</sub>O<sub>2</sub> – Me) was probably due to the presence of 9 $\beta$ ,19-cyclopropyl group rather than the double bond in the ring system [7]. The *RR*, of the steryl acetate was 2.09 (SE-30), 2.05 (Dexsil-300), 2.09



(OV-17) and 2.05 (OV-210). The data from MS, GLC and argentation TLC were in good agreement with those of authentic 4 $\alpha$ ,14 $\alpha$ ,24-trimethyl-9 $\beta$ ,19-cyclo-5 $\alpha$ -cholest-24-en-3 $\beta$ -yl acetate (**2b**, R' = Ac) prepared from cycloeucaenyl acetate (**2a**, R' = Ac). Thus, the steryl acetate B is considered to have the structure **2b**, R' = Ac.

Though isomerisation of a sterol with the  $\Delta^{24(28)}$  side chain (R = a) to a  $\Delta^{24(25)}$  side chain (R = b) during chromatography on Si gel has been recognized [8], a careful preliminary experiment using 24-methylenecycloartanol (R = a) under the same conditions used in this study showed no detectable formation of cyclobranol (24-methylcycloart-24-enol, R = b). Therefore, the two steryl acetates A and B detected in the acetylated 4-monomethylsterol fraction from the seeds of *B. napus* are considered to be natural products rather than artefacts.

There are four 24-alkylated  $\Delta^{24}$ -sterols hitherto reported to occur in higher plants: cyclobranol as its ferulate in rice-bran oil [9], 24-ethylcholesta-7,24-dien-3 $\beta$ -ol in sunflower oil [10], and 24-methylcholesta-5,24-dien-3 $\beta$ -ol and its 24-ethyl homolog in *Withania somnifera* (Solanaceae) [11]. This work appears to be the first record of the detection of 4 $\alpha$ -methyl 24-alkylated  $\Delta^{24}$ -sterols (**1b** and **2b**) in higher plants. The biogenetic importance of the 24-alkylated  $\Delta^{24}$ -sterols has been well discussed [11-16].

#### EXPERIMENTAL

Recrystallizations were performed in Me<sub>2</sub>CO-MeOH. Mp's are uncorr. IR spectra were recorded in KBr. PMR spectra were measured at 100 MHz in CDCl<sub>3</sub> with TMS as internal reference. MS (70 eV, > m/e 200) were taken with a GC-MS (2% OV-17 column) or with a probe injection. GLC analyses (cholesteryl acetate, RR<sub>s</sub> = 1.00) were carried out on 2 m  $\times$  3 mm glass columns packed with 2% SE-30 (column temp. 228°), 2% Dexil-300 (255°), 3% OV-17 (265°) and 2% OV-210 (202°) on 80-100 mesh Gas Chrom-Z, respectively, with 50 ml/min N<sub>2</sub>.

**Obtusifoliol acetate (1a, R' = Ac).** Obtusifoliol (**1a**, R' = H) was isolated from the latex of *Euphorbia regis-Jubae* W. B. as described by González *et al.* [17]. Its acetate (**1a**, R' = Ac) showed mp 106-108° (lit. [17] mp 107-108°). IR  $\nu_{\max}$  cm<sup>-1</sup>: 3080, 1638, 880 (C=CH<sub>2</sub>), 1730, 1246 (OAc). PMR  $\delta$ : 0.71 (3H, s, C-18), 0.89 (3H, s, C-32), 0.99 (3H, s, C-19), 2.05 (3H, s, 3 $\beta$ -OAc), 0.86 (3H, d, J = 6.3 Hz, C-30), 0.92 (3H, d, J = 6.9 Hz, C-21), 1.02 (6H, d, J = 7.6 Hz, C-26, 27), 4.38 (1H, m, W<sub>1/2</sub> = 27 Hz, 3 $\alpha$ -H), 4.69 (2H, bd, J = 4.4 Hz, C-28). MS (probe) m/e (rel. int.): 468 (M<sup>+</sup>, C<sub>32</sub>H<sub>52</sub>O<sub>2</sub>, 60), 453 (100), 408 (4), 393 (24), 369 (9), 341 (2), 324 (3), 301 (9), 287 (22), 227 (20).

**4 $\alpha$ ,14 $\alpha$ ,24-Trimethyl-5 $\alpha$ -cholesta-8,24-dien-3 $\beta$ -yl acetate (1b,**

R' = Ac). Isomerisation of **1a**-acetate (50 mg) with H<sub>2</sub>SO<sub>4</sub> in IPA and EtOAc under reflux [18] yielded a mixture of dienes differing in the position of the side chain double bond. Separation of this mixture by AgNO<sub>3</sub>-Si gel PLC gave **1b**-acetate (6 mg), mp 139-140°. IR  $\nu_{\max}$  cm<sup>-1</sup>: 1730, 1246 (OAc). PMR  $\delta$ : 0.70 (3H, s, C-18), 0.89 (3H, s, C-32), 0.98 (3H, s, C-19), 1.63 (9H, s, C-26, 27, 28), 2.05 (3H, s, 3 $\beta$ -OAc), 0.85 (3H, d, J = 5.9 Hz, C-30), 0.93 (3H, d, J = 6.0 Hz, C-21), 4.38 (1H, m, W<sub>1/2</sub> = 26 Hz, 3 $\alpha$ -H). MS (probe) m/e (rel. int.): 468 (M<sup>+</sup>, C<sub>32</sub>H<sub>52</sub>O<sub>2</sub>, 55), 453 (100), 408 (3), 393 (29), 384 (4), 369 (11), 341 (2), 301 (4), 287 (8), 227 (10).

**Cycloeucaenyl acetate (2a, R' = Ac).** Cycloeucaenol (**2a**, R' = H) was isolated from the unsaponifiable matter, obtained by saponification of the ferulate fraction separated from rice-bran oil, by PLC as described previously [19]. The alcohol on acetylation (Ac<sub>2</sub>O-Py) yielded the acetate (**2a**, R' = Ac), mp 108-110° (lit. [19] mp 110-111°). IR  $\nu_{\max}$  cm<sup>-1</sup>: 3080, 1638, 885 (C=CH<sub>2</sub>); 3040, 1018 (cyclopropyl), 1730, 1240 (OAc). PMR  $\delta$ : 0.90 (3H, s, C-32), 0.98 (3H, s, C-18), 2.05 (3H, s, 3 $\beta$ -OAc), 0.16 (1H, d, J = 4.2 Hz, C-19), 0.42 (1H, d, J = 4.2 Hz, C-19), 0.86 (3H, d, J = 5.7 Hz, C-30), 1.03 (6H, d, J = 6.0 Hz, C-26, 27), 4.71 (2H, bd, J = 4.3 Hz, C-28), 4.56 (1H, m, W<sub>1/2</sub> = 26 Hz, 3 $\alpha$ -H). MS (probe) m/e (rel. int.): 468 (M<sup>+</sup>, C<sub>32</sub>H<sub>52</sub>O<sub>2</sub>, 7), 453 (8), 408 (100), 393 (51), 384 (5), 365 (7), 343 (5), 300 (10), 285 (3), 283 (13).

**4 $\alpha$ ,14 $\alpha$ ,24-Trimethyl-9 $\beta$ ,19-cyclo-5 $\alpha$ -cholest-24-en-3 $\beta$ -ol (2b, R' = Ac).** Isomerisation of **2a**-acetate with H<sub>2</sub>SO<sub>4</sub> as described above yielded **2b**-acetate, mp 159-160°. IR  $\nu_{\max}$  cm<sup>-1</sup>: 3040, 1020 (cyclopropyl), 1732, 1240 (OAc). PMR  $\delta$ : 0.89 (3H, s, C-32), 0.97 (3H, s, C-18), 1.63 (9H, s, C-26, 27, 28), 2.05 (3H, s, 3 $\beta$ -OAc), 0.17 (1H, d, J = 4.4 Hz, C-19), 0.42 (1H, d, J = 4.4 Hz, C-19), 0.86 (3H, d, J = 7 Hz, C-30), 0.91 (3H, d, J = 7 Hz, C-21), 4.52 (1H, m, W<sub>1/2</sub> = 25 Hz, 3 $\alpha$ -H). MS (probe) m/e (rel. int.): 468 (M<sup>+</sup>, C<sub>32</sub>H<sub>52</sub>O<sub>2</sub>, 19), 453 (7), 408 (100), 393 (59), 384 (2), 369 (1), 353 (7), 341 (2), 309 (5), 301 (2), 287 (1), 285 (5), 227 (3).

**Acknowledgements**—We thank Dr. B. M. Fraga, University of La Laguna, Tenerife, Spain, and Riken Vitamin Oil Co., Tokyo, for generous gifts of the latex of *Euphorbia regis-Jubae* and the ferulate of rice-bran oil. Our thanks are also due to Dr. Y. Toyama for valuable comments and advice.

#### REFERENCES

- Itoh, T., Tamura, T. and Matsumoto, T. (1976) *Phytochemistry* **15**, 1781.
- Jeong, T. M., Itoh, T., Tamura, T. and Matsumoto, T. (1975) *Lipids* **10**, 634.
- Knights, B. A. and Berrie A. M. M. (1971) *Phytochemistry* **10**, 131.
- Itoh, T., Tamura, T. and Matsumoto, T. (1973) *J. Am. Oil Chem. Soc.* **50**, 300.
- Sawicki, J. and Mordret, R. (1970) *Rev. Fr. Corps Gras* **17**, 685.
- Wyllie, S. G. and Djerassi, C. (1968) *J. Org. Chem.* **33**, 305.
- Aplin, R. T. and Hornby, G. M. (1966) *J. Chem. Soc. B* 1078.
- Nair, M. G. and Chang, F. C. (1971) *Tetrahedron Letters* 2513.
- Endo, T., Misu, O. and Inaba, Y. (1969) *Yukagaku* **18**, 255.
- Homberg, E. E. and Schiller, H. P. T. (1973) *Phytochemistry* **12**, 1767.
- Lockley, W. J. S., Roberts, D. P., Rees, H. H. and Goodwin, T. W. (1974) *Tetrahedron Letters* 3773.
- Tomita, Y. and Uomiri, A. (1972) *Chem. Commun.* 1416.
- Goad, L. J. and Goodwin, T. W. (1972) *Progr. Phytochemistry* **3**, 113.
- Randall, P. J., Rees, H. H. and Goodwin, T. W. (1972) *Chem. Commun.* 1295.
- Argarego, W. L. R., Goad, L. J. and Goodwin, T. W. (1973) *Phytochemistry* **12**, 2181.

